Alterations in haematobiochemical parameters in crossbred cattle suffering from Babesiosis

Gaurav Charaya¹, Parveen Goel¹, Jasleen Kaur², Maneesh Kumar³ and Yudhbir Singh¹

¹Department of Veterinary Medicine, ²Department of Veterinary Microbiology, ³Department of Teaching Veterinary Clinical Complex, College of Veterinary Sciences, Lala Lajpat Rai University of Veterinary and Animal Sciences, Hisar, Haryana

Bovine babesiosis is an important tick-borne disease affecting animals all over the world causing huge economic losses in subtropical and tropical territories. Projected economic losses to livestock due to babesiosis in India are estimated to be about 84.7 million US dollars (Narladkar, 2018). Disease is caused by an intraerthrocytic hemoprotozoan affecting animal erythroctytes and transmitted by blood-sucking ticks of the Ixodidae family. In India, bovine babesiosis is mainly caused by *Babesia bigemina* and *Babesia bovis* transmitted by *Boophilus microplus* tick. The crossbred cattle exhibited higher rate of susceptibility than zebu and buffaloes, which mainly act as carrier.

Clinically, disease has been characterized by anemia, fever, hemoglobinuria, and death (Mosqueda *et al.*, 2012). Presence of the protozoan inside red cells is considered as confirmative especially during acute stage of the disease (Singh *et al*, 2000). Researchers in past have shown deleterious effect of parasite on the health of cattle leading to liver and kidney dysfunction (Hamoda *et al.*, 2014, Aziz *et al.*, 2020, Debbarma *et al.*, 2020). The extent of damage caused by parasite determines the severity of disease and prognosis. Therefore, the present study was undertaken with an aim to determine impact of parasite on liver and kidney functioning in animals suffering from clinical babesiosis by evaluating haematobiochemical parameters.

Blood and serum samples of 56 cross bred cattle were collected for determination of haematobiochemical alterations in cattle suffering from babesiosis. Blood samples were screened for babesiosis by examination of thin blood smears stained with Giemsa stain for 30 min. About 20 microscopic fields, per slide, were observed to view the parasite. The presence of single piroplasm was recorded as positive for particular parasite and correlated with clinical signs to determine clinically infected cattle and apparently healthy cattle.

Complete haematological examination was done using fully automated haematology cell counter (MS4s, Melet Schloesing Laboratories, France). The parameters measured were haemoglobin (Hb) in g/dl, total leucocyte count (TLC) in 10³/mm³, differential leukocyte count (DLC lymphocytes(L) in %, monocytes (M) in %, neutrophils (N) in %, eosinophils (E) in % and basophils (B) in %. The serum samples were analyzed for estimation of biochemical test profile by using fully automated random access clinical chemistry analyzer (EM Destiny 200, Erba Diagnostics Mannheim GmbH-Germany). The serum biochemical parameters measured were aspartate aminotransferase in U/L, total protein in g/dl, albumin in g/dl, bilirubin direct (BID) in mg/dl, bilirubin total (BIT), blood urea nitrogen in mg/dl, creatinine in mg/ dl, calcium (Ca) in mg/dl and phosphorus (P) in mg/ dl. Independent t test was applied for determination of mean values and to compare positive sample values with negative samples values.

The mean values of haematological and biochemical parameters in nine cattle suffering from babesiosis and their comparison with 15 apparently healthy animals have been shown in Table no 1 and Table no 2. Severe anemia was recorded in cattle suffering from babesiosis with significantly lower (P<0.05) mean values of Hb, TEC and PCV in infected animals. Results obtained in present study were in agreement with results of Abdel-Hamied et al. (2020) and Aziz et al. (2020) showing severe anaemia in animals suffering from babesiosis. This severe anaemia can be attributed to massive intravascular hemolysis associated with presence Babesia spp. inside RBCs (Callow and Pepper, 1974), production of autoantibodies directed against circulating erythrocytes (Goes et al., 2007) and increased phagocytosis of parasitized and even unaffected erythrocytes by activated macrophages (Court et al., 2001).

Differential leukocyte count value showed significant decrease in neutrophil count and significant increase in lymphocyte count in infected animals as

Corresponding author: gcharaya9@gmail.com

Charaya et al.

| Parameters | Infected cross bred cattle (n=9) | Non infected cross bred cattle (15) |
|---------------------------|----------------------------------|-------------------------------------|
| Hb (g/dl) | 5.06+0.716* | 9.55±0.3075 |
| TEC (10 ⁶ /dl) | 2.58±.0.32* | 6.52±.0.19 |
| PCV (%) | 18.5±2.58* | 31.30±0.84 |
| TLC (10 ³ /ml) | 6.97 ± 1.55 | 5.54±0.46 |
| Neutrophil (%) | 48.86±9.04* | $63.28{\pm}2.05$ |
| Lymphocyte (%) | 46.06±9.31* | 32.38±1.96 |
| Basophil (%) | 0.5+.3521 | 0.456+.2828 |
| Eosinophil (%) | 2.95±0.93 | 6.82±2.33 |
| Monocyte (%) | 5.02±1.0 | 4.20±0.35 |

Table 1. Haematological alterations in cross bred cattle suffering from babesiosis

*significant at P< 0.05

compared to apparently healthy animals. This finding was in accordance with the results of Tufani *et al.* (2015) and Abdel-Hamied *et al.* (2020). Reasons for increase in lymphocyte can be stimulation of phagocytic cells like lymphocytes and monocytes associated with RBCs breakdown for removal of the toxic remnants of damaged erythrocytes (Guglielmone *et al.*, 1996) and activation of body defense mechanisms for antibodies production against the protozoan in response to babesia infection (Court *et al.*, 2001). The significant reduction in neutrophilic sequestration in the spleen, haematopoietic precursor, cell damage, increased neutrophil adherence, or a combination of all (Akel and Mobarakai, 2017).

Significant elevated level of AST was found as compared to level observed in apparently healthy cattle. AST enzyme levels are the indicators of hepatic function and the rise in serum AST values may be due to alteration of liver function as a result of bovine babesiosis (Zulfiqar et al., 2012). Mean values of urea and creatinine was found to be significantly increased (P<0.05) in cattle suffering from clinical babesiosis clearly indicating renal function impairment in infected cattle. This can be attributed to hypoxic and toxic renal tissue damage as a result of hypoxia, hemoglobinuria and increased catabolism of haemoglobulin (Hamoda et al., 2014). There were significant elevation in the serum bilirubin levels (direct and total bilirubin) that was attributed to the intense hemolysis and hepatic dysfunction in diseased animals (Schwint et al., 2009). Significant reduction (P<0.05) in total protein values and non significant decrease in albumin was observed in cattle suffering from babesiosis suggests parasite deteriorates the liver and kidney function drastically. These results were consistent with the findings of Shinde et al.(2019) and Aziz et al.(2020). Therefore, it can be concluded that aggressive therapy should be initiated to combat parasitemia and limit the damage of parasite to vital organs.

| Table 2. Biochemical Alterations in cross bre | d cattle suffering from | babesiosis |
|---|-------------------------|------------|
|---|-------------------------|------------|

| Parameters | Infected cross bred cattle (n=9) | Non infected cross bred cattle(n=15) |
|------------------------|----------------------------------|--------------------------------------|
| AST (U/L) | 140.93±26.10* | 58.39±3.24 |
| Phosphorous (mg%) | 6.24±0.37 | $5.82{\pm}0.35$ |
| Total Bilirubin (mg%) | 1.55 ± 1.18 | 0.10±0.013 |
| Direct Bilirubin (mg%) | $0.49{\pm}0.27$ | $0.149{\pm}0.007$ |
| Urea (mg%) | 50.014±12.06* | 9.87±1.72 |
| Creatinine (mg%) | 0.89±0.25* | $0.35 {\pm} 0.17$ |
| Calcium (mg%) | 9.00±0.56 | 11.94±0.34 |
| Protein (g%) | 5.75±0.46* | $8.42{\pm}0.29$ |
| Albumin(g%) | 2.32±0.26 | 2.43±0.11 |

*significant at P< 0.05

Acknowledgement

Authors are thankful to the Professor & Head, Veterinary Medicine & Professor & Head, Teaching Veterinary Clinical Complex for providing facilities for conduction of experiments.

References

- Akel, T. and Mobarakai, N. 2017. Hematologic manifestations of babesiosis. Ann. Clin. Microbiol. 16(6): 1-7.
- Aziz, P.R., Marodia, S., Ganesan, P.I., and Sharma, C.S. 2020. A clinical study on hemato-biochemical changes in cows affected with Babesiosis. *Pharm. Innov. J*, 9(2): 242-45.
- Callow, L. L., and Pepper, P. R. 1974. Measurement of and correlation between fever, changes in the packed cell volume and parasitemia in the evaluation of the susceptibility of cattle to infection with *Babesia argentina*. Aust. Vet. J. 50: 1-5.
- Court, R. A., Jackson, L. A. and Lee, R. P. 2001. Elevated anti-parasitic activity in peripheral blood monocytes and neutrophils of cattle infected with *Babesia bovis*. *Int. J. Parasitol.*:31: 29–37.
- Debbarma, A., Pandit, S., Jas, R., Baidya, S., Batabyal, S. and Bachan, M. 2020. Alterations of Serum Biochemical Parameter in Cattle Naturally Infected with Tick-borne Haemoparasitic Diseases in West Bengal, India. *Int. J. Livest. Res.* 10 (9): 91-95.
- Goes, T. S., Goes, V. S., Ribeiro, M. F. and Gontijo, C. M. 2007. Bovine babesiosis: anti-erythrocyte antibodies purification from the sera of naturally infected cattle. *Vet. Immunol. Immunopathol.* 116: 215–18.
- Guglielmone, A. A., Gaido, A. B. and Mangold, A. J. 1996. Light microscopy diagnosis of Babesia bovis and Babesia bigemina kinetes in the hemolymph of artificially infected Boophilus microplus engorged female ticks. *Vet. Parasitol.* 61(1-2): 15-20.

- Hamoda, A. F., Radwan, M., Rashed, R. and Amin, A. 2014. Toxic effect of babesiosis in cattle and chemotherapiotic treatment in Egypt. Am. J. Infect. Dis. Micro. 2: 91-96.
- Khinchi, R. K., Bihani, D. K., Ahuja, A. and Singh, A. P. 2016. Haemato-biochemical changes in Babesia infected cattle. *Vet. Pract.* 17(1): 59-60.
- Narladkar, B. W. 2018. Projected economic losses due to vector and vector-borne parasitic diseases in livestock of India and its significance in implementing the concept of integrated practices for vector management. *Vet. World.* 11(2):151-60.
- Mosqueda, J., Olvera-Ramirez, A., Aguilar-Tipacamu, G. and Canto, G.J. 2012. Current advances in detection and treatment of babesiosis. *Curr. Med. Chem.* **19**:1504–18.
- Schwint, O. N., Ueti, M. W., Palmer, G. H., Kappmeyer, L. S., Hines, M. T., Cordes, R. T., Knowles, D. P. and Scoles, G. A. 2009. Imidocarb dipropionate clears persistent *Babesia caballi* infection with elimination of transmission potential. *Antimicrob. Agents Chemother.* 53: 4327-32.
- Shinde, R. M., Bhikane, A. U., Naraladkar, B. W. and Masare, P. S. 2019. Antioxidants as an adjunct therapy in clinical management of babesiosis in cattle: A novel approach. *Ruminant Sci.* 8(1): 93-100.
- Singh, A.P., Singla, L.D. and Singh, A. 2000. A study on the effects of macroclimatic factors on the seasonal population dynamics of *Boophilus micropus* (Canes 1888) infesting the cross-bred cattle of Ludhiana district. *Int J Anim Sci.* 15: 29–31.
- Tufani, N. A., Fazili, M. R., Malik, H. U., Beigh, S. A. and Dar, K. H. 2015. Clinico Haematological Profile and Therapeutic Management of Acute Babesiosis in a Holstein-Friesian Crossbred Cow. Vet. Clin.Sci. 3(3): 1-14.
- Zulfiqar, S., Shahnawaz, S., Ali, M., Bhutta, A. M., Iqbal, S., Hayat, S., Qadir, S., Latif M., Kiran, N., Saeed, A., Ali, M. and Iqbal, F. 2012.Detection of *Babesia bovis* in blood samples and its effect on the hematological and serum biochemical profile in large ruminants from Southern Punjab. *Asian Pac. J. Trop. Biomed.* 2(2): 104-08.

Received : 10.12.2020 Accepted : 09.06.2021